

Exploring the Matrix of Organisms in which Invasive Plants Thrive

- Parker and Swope
 - Plant-microbe interactions and plant invasions.
- Wojcik
 - Native pollinators and invasive plants: Implications for agriculture and restoration.
- Aslan
 - Mutualisms between native and non-native species: Global trends and Californian case studies.

Plant-Microbe Interactions and Plant Invasions

Ingrid Parker

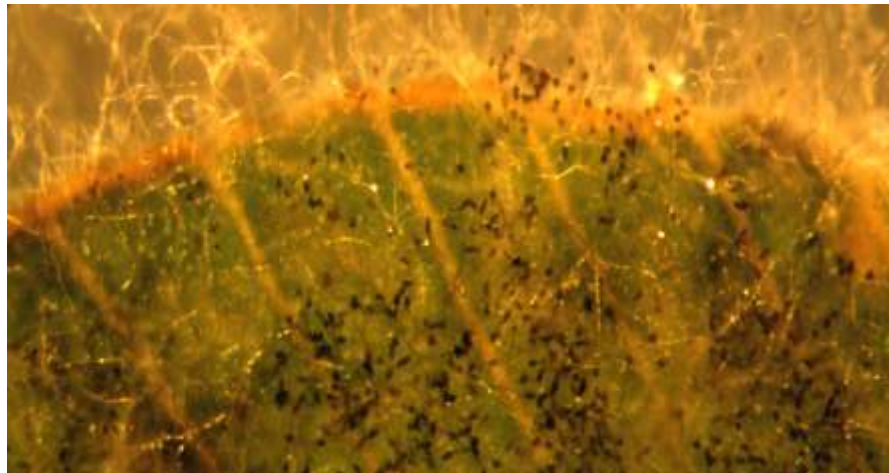
UC Santa Cruz

Sarah Swope

USDA Exotic & Invasive Weeds Research Unit

Gregory Gilbert

UC Santa Cruz



Microbes are everywhere



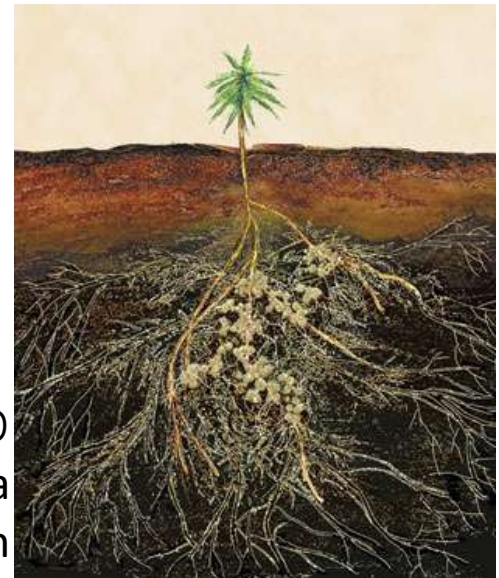
Southern corn leaf
blight



Potato blight



Sudden oak death



Mycorrhizae!

©
Melissa
Buntin

Pathogen



Host



Native

Novel

Native

**[Native
interaction]**

**Novel
epidemic**

Novel

**Introduced
plant**

**Biological
control**

Pathogen



Native

Novel

Host



Native

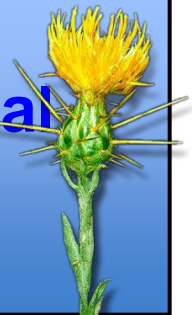
Novel

**[Native
interaction]**

**Novel
epidemic**

**Introduced
plant**

**Biological
control**



Do introduced species always “leave natural enemies behind”?

- Do pathogens explain or constrain invasion success?

ECOLOGY

- How will these interactions change over time?

EVOLUTION



A photograph of a coastal landscape. In the foreground, a person wearing a blue jacket, a dark hat, and sunglasses is sitting on a hillside covered in green vegetation with small yellow and purple flowers. The person is holding a camera. In the background, there is a body of water with waves crashing against a rocky shore. A large, multi-story building with a red roof is situated on a grassy hill overlooking the water. The sky is clear and blue.

Trifolium/Medicago

9 native species

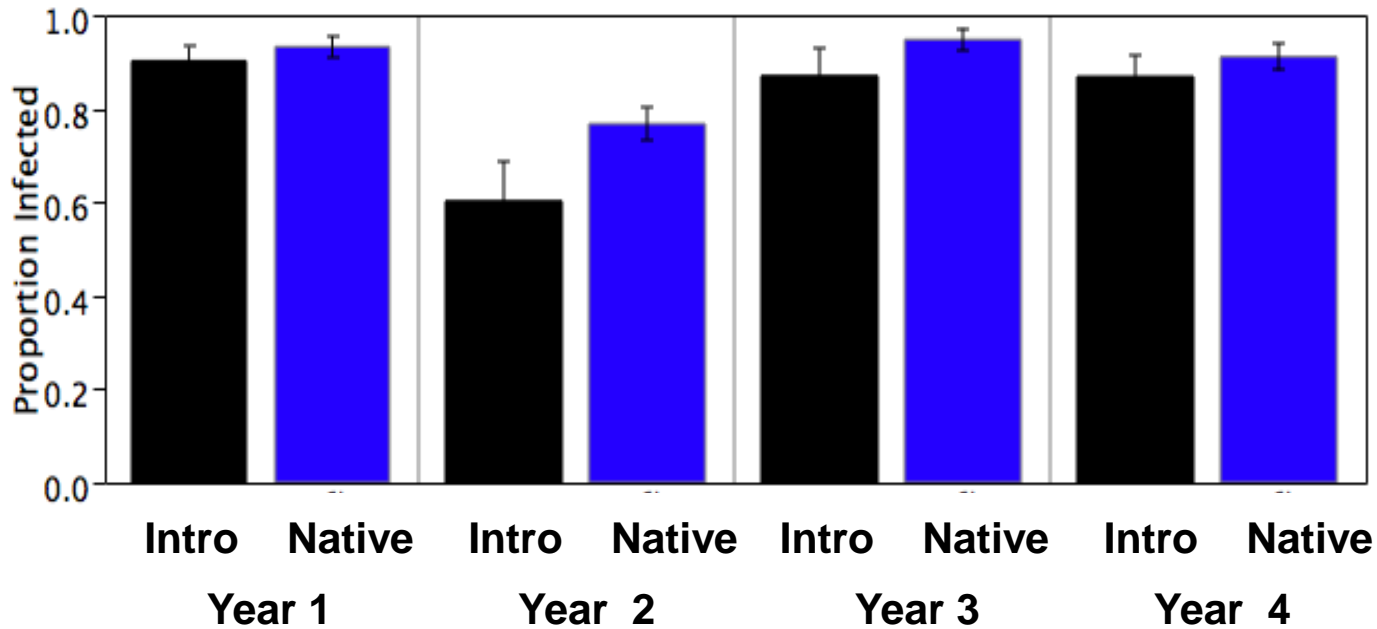
8 introduced species

[species as replicate]

Greg Gilbert, UCSC



FUNGAL INFECTION

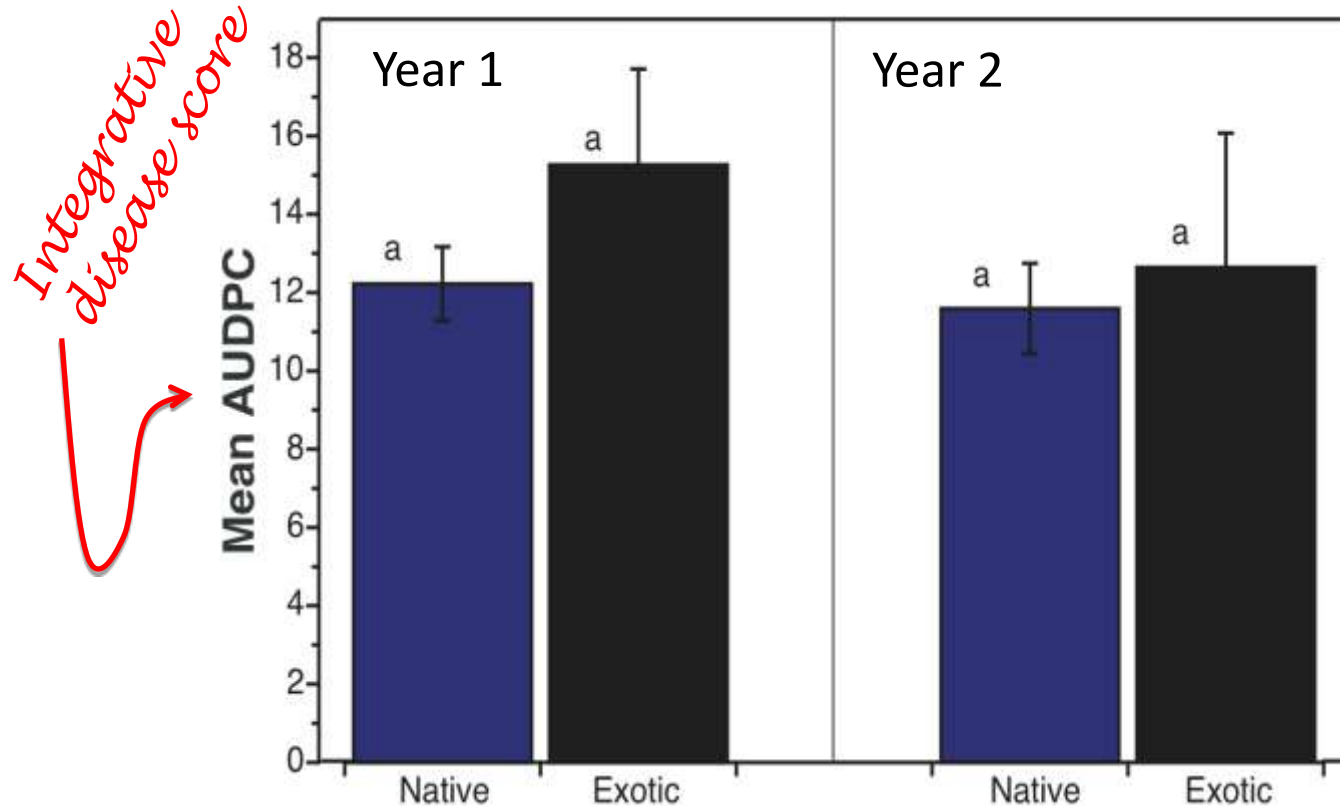


Infection is extremely common

Introduced \approx Native



DISEASE PREVALENCE



Introduced \approx Native
Same for disease severity.
Same for fitness cost.





© Forest & Kim Starr

Vs.



© trnerr.org

NON-INVASIVE

T. glomeratum

T. campestre

T. Subterraneum

T. repens

T. dubium

M. lupulina

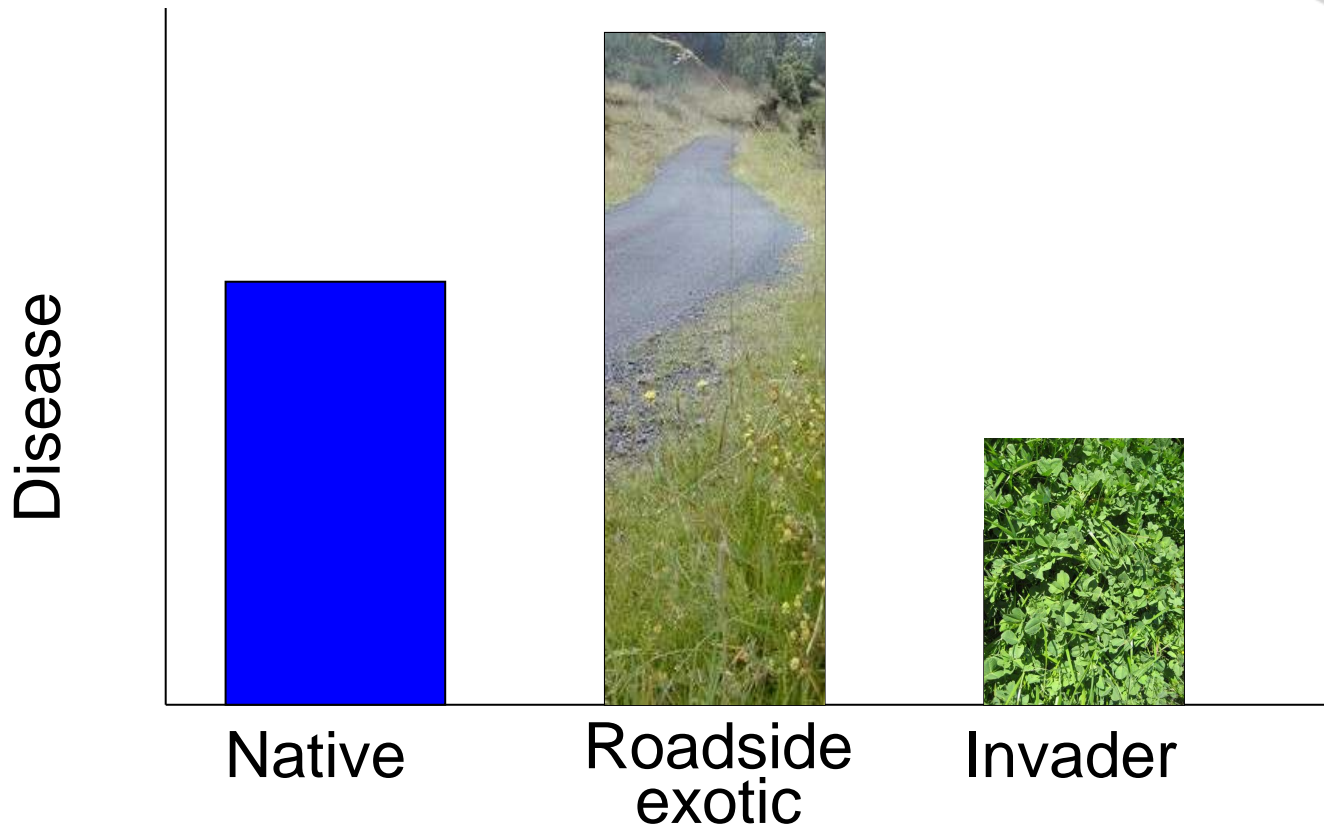
INVASIVE

M. polymorpha

M. arabica

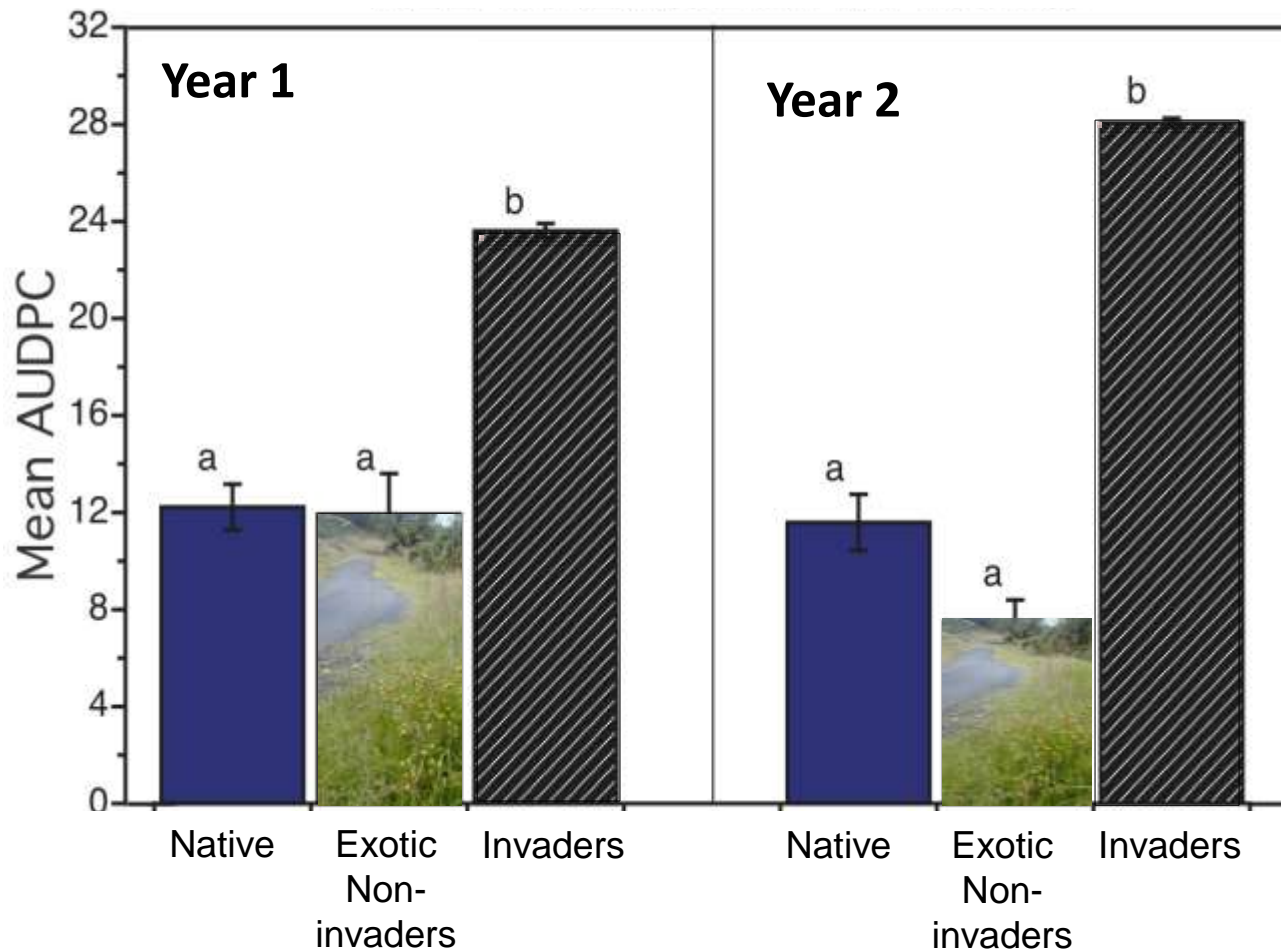
Does BIOTIC RESISTANCE limit some introduced clovers to roadsides?

Predicted
Results



NO:
The more invasive species
experienced MORE disease.

Actual
Results



CONCLUSIONS:

Clovers and Their Pathogens



Introduced & native plants experienced similar rates of infection by pathogens.

Introduced & native plants suffered similar disease.

Of the introduced plants, the more invasive ones experienced more disease, not less.

No support for “escape from enemies”



WHY

No support for “escape from pathogens”?

Bodega clover pathogens are dominated by:

- Host-generalists (> 95% of 1178 fungi cultured)

FUNGUS RECORDED	Native hosts										Introduced hosts						
	T. barbigerrum	T. bifidum	T. fucatum	T. gracilentum	T. macraei	T. microcephalum	T. microdon	T. willdenovii	T. wormskjoldii	M. arabica	M. lupulina	M. polymorpha	T. campestre	T. dubium	T. glomeratum	T. repens	T. subterraneum
Fungus																	
<i>Stemphylium</i> spp.																	
<i>Alternaria</i> sp.																	
<i>Cladosporium</i> sp.																	
<i>Colletotrichum trifolii</i>																	
<i>Phoma</i> sp.																	
<i>Cochliobolus spicifer</i>																	
<i>Cylindrocarpon</i> sp.																	
<i>Leptosphaerulina trifolii</i>																	
<i>Uromyces minor</i>																	
<i>Pseudopeziza trifolii</i>																	
<i>Epicoccum nigrum</i>																	
<i>Trichurus</i> sp.																	
<i>Fusarium</i> cf. <i>roseum</i>																	
<i>Chaetopsina fulva</i>																	
<i>Idriella</i> sp.																	
<i>Itersonilia</i> sp.																	
<i>Acrodictys</i> sp.																	
<i>Volutella</i> sp.																	
<i>Uromyces striatus</i>																	
<i>Ulocladium atrum</i>																	
<i>Cercospora</i> sp.																	
<i>Arthrinium</i> sp.																	

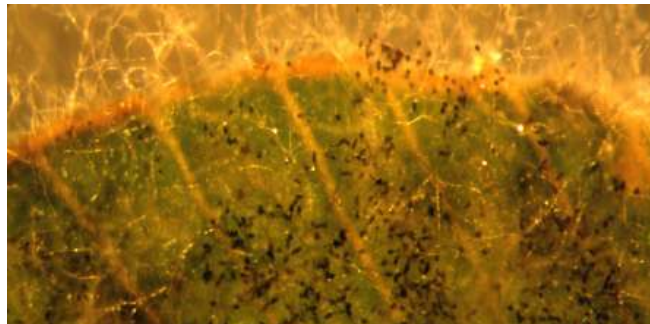


WHY

No support for “escape from pathogens”?

Bodega clover pathogens are dominated by:

- Host-generalists (> 95% of 1178 fungi cultured)
 - Taxa that have pathogenic conspecifics or congeners in the native range
-
- **Evolution of native pathogens to utilize non-native hosts?**



Evolution of native pathogens to utilize non-native hosts?



In 5 generations in the lab:

Infection rates ↑↑

CA genotypes compared to
EU genotypes:

Infection rates ↑

VIRULENCE ↓↓

Rapid evolution
will play a role in modifying interactions
between invaders & natural enemies
over time.





Sarah
Swope
USDA-ARS

Pathogen



Native

Novel



Host

Native

[Native
interaction]

Novel
epidemic

Novel

Introduced
plant

Biological
control



MICROBES FOR BIOCONTROL:

Puccinia jaceae on *Centaurea solstitialis*



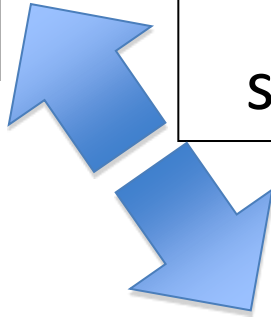
- Introduced 2004
- How will impact depend on the presence of other species?
- How will impact depend on abiotic environment?





Puccinia jaceae f.s.
solstitialis

Pathogen and seed predators
spatially & temporally
separated from one another



Eustenopus villosus

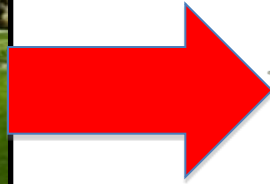


Chaetorellia succinea
(& *C. australis*)

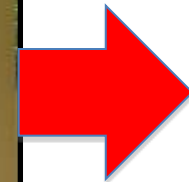


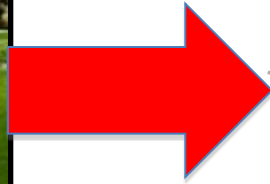
Established biocontrol insects: predispersal
seed predators





Pathogen has a
DIRECT NEGATIVE impact
on the plant (biomass,
inflorescences)

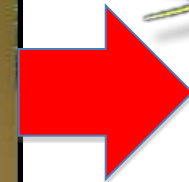




Pathogen has a
DIRECT NEGATIVE impact
on the plant (biomass,
inflorescences)

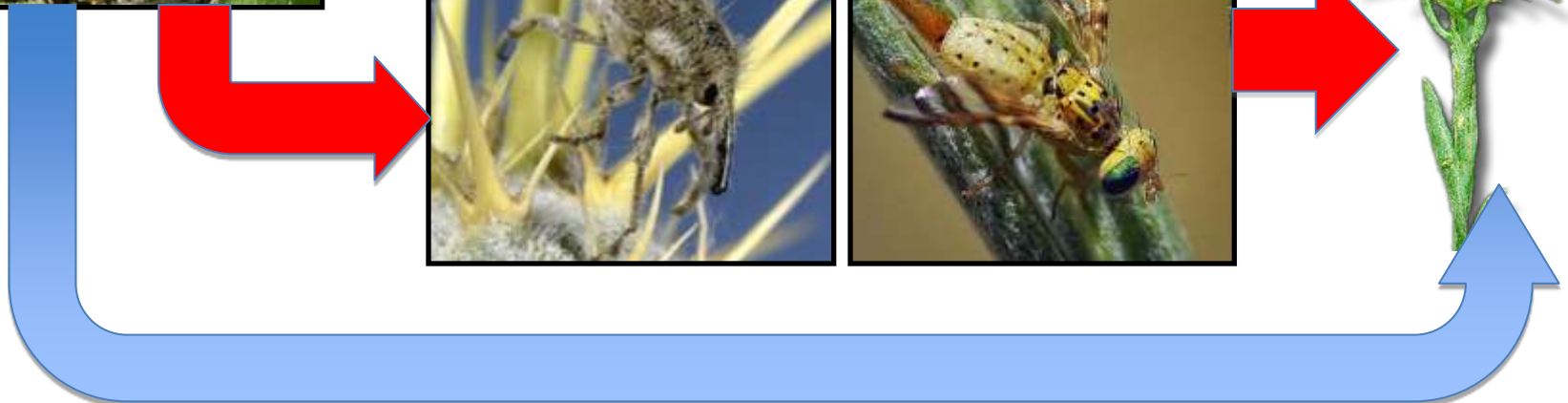


INDIRECT POSITIVE impact
on the plant through its seed predators



MECHANISM for interaction?

- Change in nutrient status?
- SAR response protects the plant against herbivores?



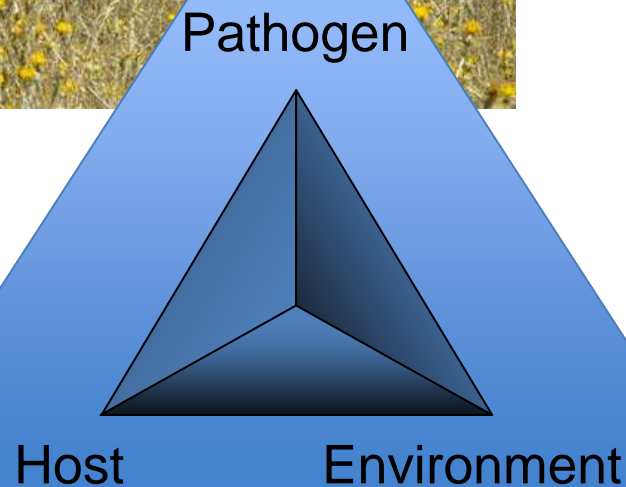
MICROBES FOR BIOCONTROL:

Puccinia jaceae on *Centaurea solstitialis*



- How will impact depend on the presence of other species?
- How will impact depend on abiotic environment?

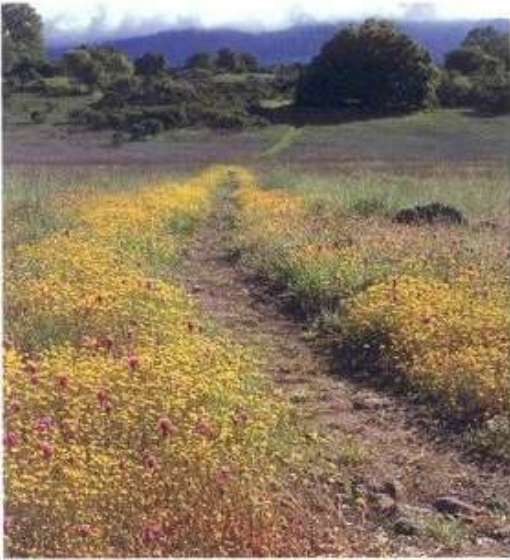
**“DISEASE
TRIANGLE”**



INTRODUCTION TO CALIFORNIA
SOILS AND PLANTS

Serpentine, Vernal Pools, and Other Geobotanical Wonders

ARTHUR R. KRUCKEBERG



Serpentine =
high conservation value

Unique properties: low $\text{Ca}^{++}/\text{Mg}^{++}$
can be stressful for plants

Ca^{++} role in
response to
infection



Is biocontrol more effective
on serpentine?

Does interaction between
agents change on serpentine?



Ilana Stein

Adjacent
patches of
serpentine and
non-serpentine
(McLaughlin NR)

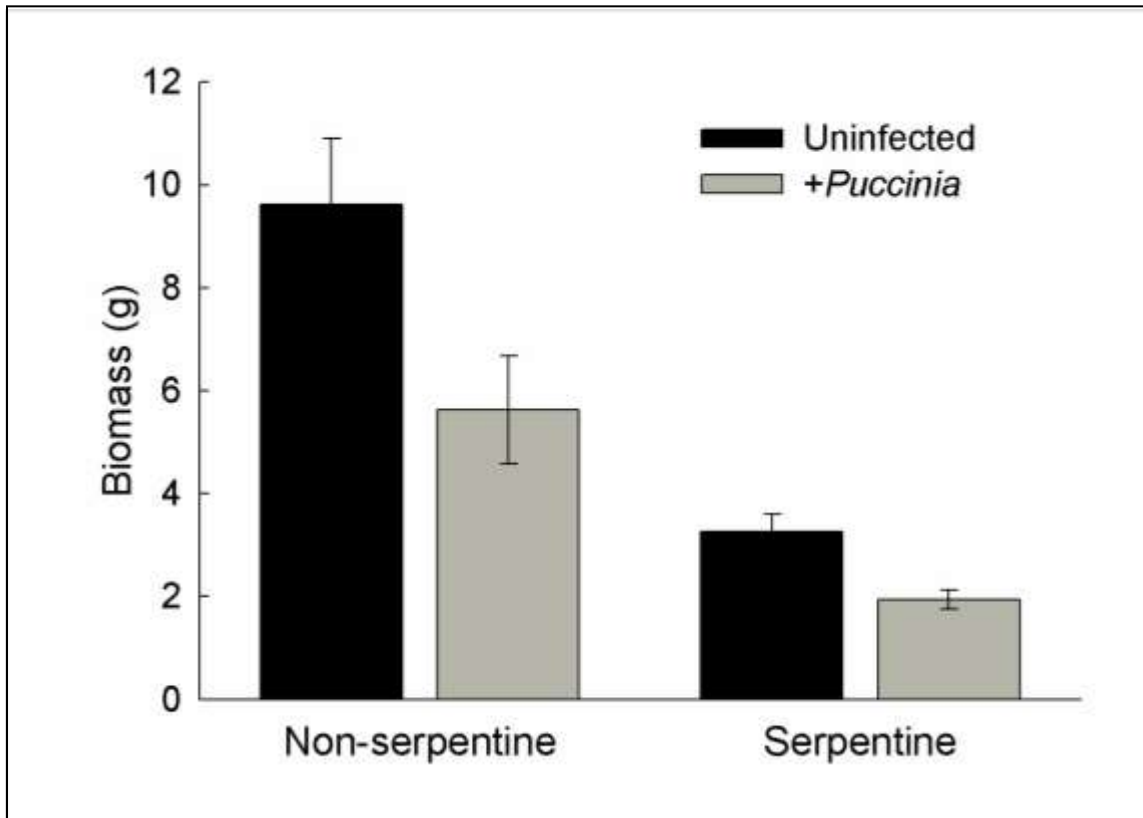


N = 400 naturally recruiting seedlings
+/- *Puccinia* inoculation

Exposed to natural levels of
attack by the seed predators



Direct impact of pathogen on plant:



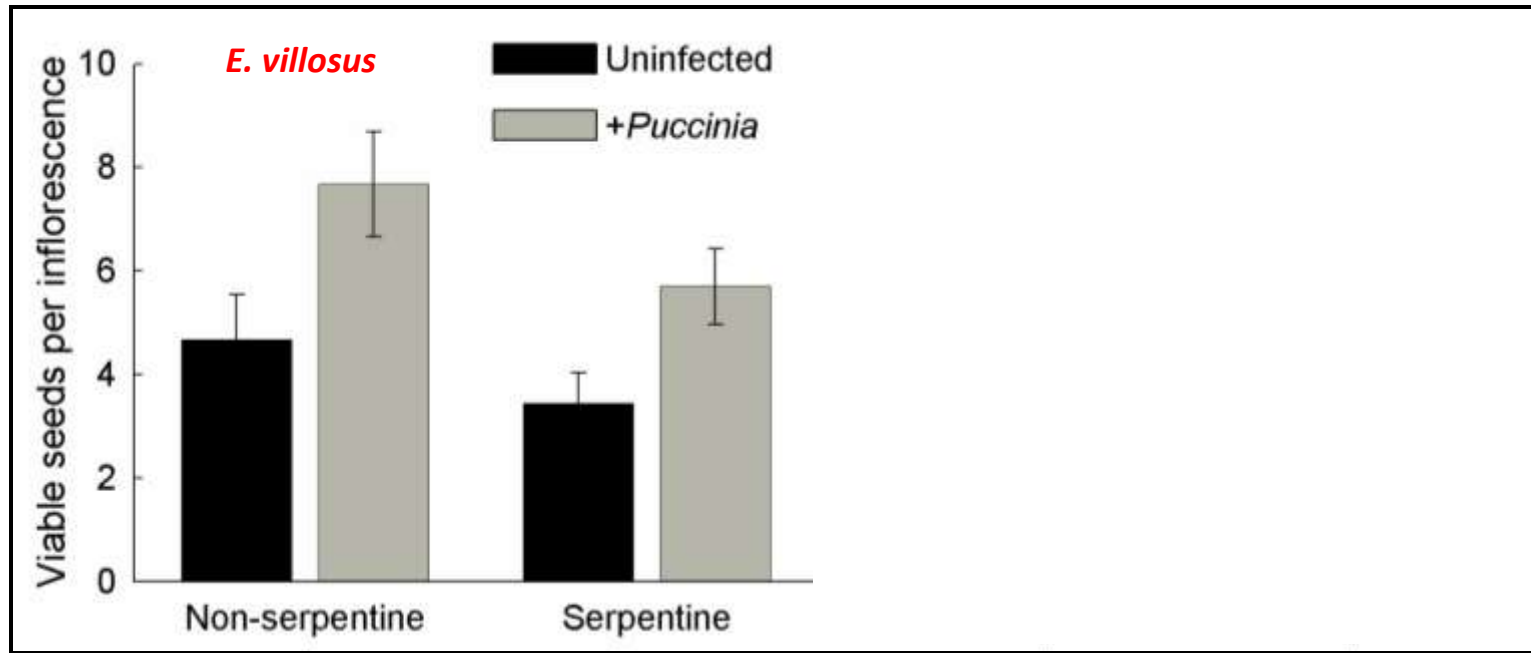
Soil type: $p=0.0001$
Pathogen: $p=0.0001$
Soil \times Pathogen: $p=0.57$

Same pattern for infl #
No effect on # viable seeds / inflorescence



Indirect impact of pathogen via seed predators:

Data for attacked inflorescences



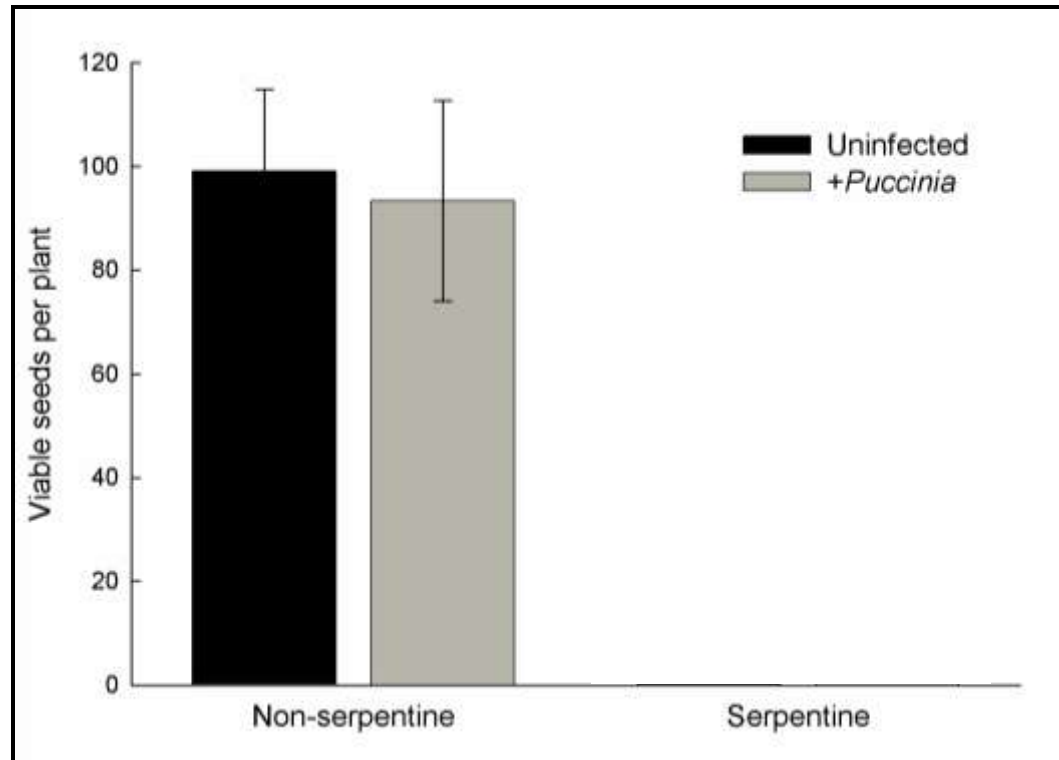
Soil type: $p=0.04$
Pathogen: $p=0.0001$
Soil \times Pathogen: $p=0.78$



Soil type: $p=0.0001$
Pathogen: $p=0.0001$
Soil \times Pathogen: $p=0.01$



Net impact of direct and indirect interactions:



Soil type: $p=0.0001$

Pathogen: $p=0.005$

Soil \times Pathogen: $p=0.009$

Non-serp soil: pathogen strongly protects the plant from seed predation = No net increase in control.

Serpentine: Adding pathogen increases control.



CONCLUSIONS:

- Microbial biological control agents can be effective...at least in some environments
- Plant-mediated interactions between biocontrol agents may affect success
- Impact of a biocontrol agent may depend on other key interactors ...
and may depend on abiotic conditions



THANKS



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Beth Howard, John Hein, and many field
assistants



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Jill Barrett Scholarship to I.S.